



**Transactions of the American Fisheries Society** 

ISSN: 0002-8487 (Print) 1548-8659 (Online) Journal homepage: http://www.tandfonline.com/loi/utaf20

# Feeding Ecology of Dolphinfish in the Western Gulf of Mexico

Rachel A. Brewton, Matthew J. Ajemian, Peter C. Young & Gregory W. Stunz

To cite this article: Rachel A. Brewton, Matthew J. Ajemian, Peter C. Young & Gregory W. Stunz (2016) Feeding Ecology of Dolphinfish in the Western Gulf of Mexico, Transactions of the American Fisheries Society, 145:4, 839-853, DOI: 10.1080/00028487.2016.1159614

To link to this article: http://dx.doi.org/10.1080/00028487.2016.1159614

	1	1	(	1
1				
1				
-1				

Published online: 24 Jun 2016.



Submit your article to this journal 🗹

Article views: 36



View related articles 🗹



View Crossmark data 🗹

Full Terms & Conditions of access and use can be found at http://www.tandfonline.com/action/journalInformation?journalCode=utaf20

## ARTICLE

# Feeding Ecology of Dolphinfish in the Western Gulf of Mexico

#### **Rachel A. Brewton**

Harte Research Institute for Gulf of Mexico Studies, Texas A&M University–Corpus Christi, 6300 Ocean Drive, Corpus Christi, Texas 78412, USA

#### Matthew J. Ajemian

Florida Atlantic University, Harbor Branch Oceanographic Institute, 5600 U.S. Highway 1 North, Fort Pierce, Florida 34946, USA

### Peter C. Young and Gregory W. Stunz\*

Harte Research Institute for Gulf of Mexico Studies, Texas A&M University–Corpus Christi, 6300 Ocean Drive, Corpus Christi, Texas 78412, USA

#### Abstract

Dolphinfish Coryphaena hippurus support important commercial and recreational fisheries in the Gulf of Mexico. Understanding the feeding ecology of this economically important pelagic fish is key to its sustainable management; however, dietary data from this region are sparse. We conducted a comprehensive diet study to develop new trophic baselines and investigate potential ontogenetic and sex-related shifts in Dolphinfish feeding ecology. The stomach contents of 357 Dolphinfish (27.6-148.5 cm TL) were visually examined from fishery-dependent sources off Port Aransas, Texas. Our analyses revealed a highly piscivorous diet with Actinopterygii comprising 70.44% of the stomach contents by number. The most commonly observed taxa were carangid (12.45% N) and tetraodontiform (12.08%N; families Balistidae, Monacanthidae, and Tetraodontidae) fishes. Malacostracans were also common (24.83%N), mostly in the form of pelagic megalopae. Other prey categories included squid and the critically endangered Kemp's Ridley sea turtles Lepidochelys kempii. Although increasingly commom in larger fish, Sargassum spp. was found across a range of sizes in Dolphinfish, indicating that these fish feed from this drifting macroalgae throughout ontogeny. An ontogenetic shift from primary consumption of carangids and brachyurans in smaller size-classes to tetraodontids, monocanthids, and squid in larger size-classes was also observed. No sex-related difference in diet was observed. The overall infection rate for gastric parasites was 54%, an order of magnitude increase from that previously reported in the region, but consistent with recent studies from other areas. Trematode parasites in the genus Dinurus were found in 55% of stomachs and nematode parasites of the family Raphidascarididae in 16% of stomachs. Overall, Dolphinfish in the western Gulf of Mexico are highly opportunistic carnivores with a gastrointestinal parasite burden consistent with that reported in other ocean basins. These data will be important for sustainable management of this economically important species.

Numerous studies have demonstrated the key role of top predatory fishes in structuring marine communities (Bascompte et al. 2005; Frank et al. 2005; Heithaus et al. 2008; Baum and Worm 2009; Shackell et al. 2010). However, for logistical reasons, many of these studies have focused on coastal or

nearshore ecosystems, leaving a major gap in our understanding of how offshore species regulate food web dynamics in the more remote pelagic habitats. Such data limitations impede management of these economically important pelagic fish species and confound predicting potential ecosystemwide effects of

<sup>\*</sup>Corresponding author: greg.stunz@tamucc.edu

Received November 20, 2015; accepted February 17, 2016

removing these predators from pelagic ecosystems. Thus, more diet information on pelagic predators is needed to understand trophic interactions in these poorly studied environments.

The Dolphinfish Coryphaena hippurus is an epipelagic fish with circumglobal distribution in tropical to subtropical waters (Gibbs and Collette 1959) and represents a model species in which feeding habits of pelagic predators can be examined. This species is fast growing and short lived, with an average life span of approximately 3–4 years (Schwenke and Buckel 2008). Dolphinfish reach sexual maturity at around 50 cm TL (Massuti and Morales-Nin 1997; Schwenke and Buckel 2008) which correlates to an age of approximately 6 months for fish in the western Gulf of Mexico (Young 2014). Dolphinfish grow rapidly in the first year, reaching lengths from 80 to 150 cm TL; afterwards growth slows, and an estimated maximum of 160-200 cm TL is reached by approximately age 3 (Palko et al. 1982; Lasso and Zapata 1999; Schwenke and Buckel 2008; Young 2014). Dolphinfish in the Gulf of Mexico and the Caribbean Sea tend to have faster growth rates than those in other regions (Schwenke and Buckel 2008). However, the cause of this accelerated development and its relation to diet is unknown.

Perhaps due to their fast-growing nature, Dolphinfish are known to be voracious eaters with a high metabolic rate (Benetti et al. 1995). The diet of Dolphinfish has been examined in various ocean basins, including the Mediterranean Sea (Bannister 1976; Massuti et al. 1998), Arabian Sea (Varghese et al. 2013), Caribbean Sea (Oxenford and Hunte 1999), western Atlantic Ocean (Manooch et al. 1984; Rudershausen et al. 2010), and eastern Pacific Ocean (Olson and Galván-Magaña 2002; Torres-Rojas et al. 2014; Tripp-Valdez et al. 2015). However, only one study has examined Dolphinfish diets in the Gulf of Mexico (Manooch et al. 1984), and this was conducted several decades ago. Thus, there is a lack of recent. region-specific, dietary information. However, Dolphinfish are piscivorous predators with a diverse diet (Gibbs and Collette 1959; Manooch et al. 1984; Massuti et al. 1998; Oxenford and Hunte 1999; Castriota et al. 2007; Rudershausen et al. 2010; Varghese et al. 2013). In total, over 34 families of fish have been reported in the Dolphinfish diet (Manooch et al. 1984; Varghese et al. 2013).

Dolphinfish are primarily surface feeders that associate with floating offshore objects such as fish-aggregating devices (Castriota et al. 2007; Taquet et al. 2007) or floating beds of *Sargassum* spp. (Manooch et al. 1984; Rudershausen et al. 2010). Fish size can influence the foraging strategy of Dolphinfish and in some areas larger size-classes may not rely on flotsam for food resources. Specifically, larger, older fish appear to display more complex behaviors when foraging, such as swimming in circles or surfing on waves while hunting elusive prey, while smaller and younger Dolphinfish are more reliant on prey associated with flotsam (Nunes et al. 2015). Additionally, sex-related differences in feeding behavior have been observed. For example, in some regions such as the western Atlantic Ocean and the Caribbean Sea, males tend to consume more open-water species than do females that consume more flotsam-associated prey (Rose and Hassler 1974; Oxenford and Hunte 1999). Conversely, in the eastern Pacific Ocean, there is apparently no difference in the diet between sexes (Castriota et al. 2007; Tripp-Valdez et al. 2010). To date, there is little understanding of dietary or trophic differences with ontogeny or sex in the western Gulf of Mexico.

High species richness and a high intensity of parasite infection are common in epipelagic fishes (Marcogliese 2002). Dolphinfish are host to a diverse and unique community of gastric parasites (Burnett-Herkes 1974; Raptopoulou and Lambertsen 1987; Dyer et al. 1997; Carbonell et al. 1999; Williams and Bunkley-Williams 2009). Specifically, Dolphinfish are the definitive host for digenean parasites of the genus Dinurus, which are commonly found in large numbers in the stomach cavity (Carbonell et al. 1999; Williams and Bunkley-Williams 2009). However, Dinurus parasites are not considered to be significant pathogens that would negatively impact the host's quality of life (Raptopoulou and Lambertsen 1987), but they are common endoparasites of Dolphinfish in both the Mediterranean Sea and the western Atlantic Ocean (Raptopoulou and Lambertsen 1987; Carbonell et al. 1999), which demonstrates the potential connectivity between these distant habitats. Thus, the presence of certain parasites in stomachs can act as biological tags and yield information on fish migration and feeding ecology (Carbonell et al. 1999). Presently, the characterization of gastric parasites of Dolphinfish has been limited in the Gulf of Mexico, impeding our understanding of potential infection rates and interbasin connectivity.

In the western Gulf of Mexico, Dolphinfish support economically important recreational and commercial fisheries (Thompson 1999), but the dietary resources enabling Dolphinfish to sustain their relatively high growth rates are not well studied. Such data have become very relevant in this region, where the oil spill from the Deepwater Horizon in 2010 exposed early life stages of several pelagic predatory fishes, including Dolphinfish, to crude oil and dispersants (Rooker et al. 2013). Exposure of larval and juvenile fish to these toxicants may have impaired swimming performance (Mager et al. 2014), which has implications for increased mortality. Given their great capacity for large-scale migration (Merten et al. 2014a), Dolphinfish have a high potential to integrate contaminants across the Gulf of Mexico. These findings suggest a need for more information on the feeding biology of these fish in the Gulf of Mexico, as this stock is potentially recovering from a poor 2010 cohort (Kitchens and Rooker 2014).

Our overall goal was to develop new baselines of Dolphinfish feeding ecology in the northwestern Gulf of Mexico region and provide data to support the sustainable management of this species. Specifically, we sought to produce a specific survey of diets for Dolphinfish in the western Gulf of Mexico and quantify their prey in relation to ontogeny and sex. Further, a preponderance of gastric parasites in stomach contents provided us with an additional opportunity to assess the prevalence of infection in Dolphinfish.

#### **METHODS**

Dolphinfish (n = 357) were collected from fish processing houses in Port Aransas, Texas, from October 2010 to December 2011. All fish were captured in Gulf of Mexico waters near Port Aransas. Fish were measured for FL and TL to the nearest centimeter. Fish were sexed by visual inspecting external morphology (Beardsley 1967) and dissected reproductive organs when available. Due to the fishery-dependent nature of the sampling, collection dates were unavailable for some fish. Whole weight (WW) was directly obtained for 32 individuals, while estimates for the remaining individuals were calculated using FL–WW conversions following that of Young (2014):

$$WW = 1.5696 - 0.0915 \times FL + 0.0017 \times FL^2$$
.

Stomachs were removed from individuals, placed intact into perforated plastic bags, and fixed in 10% formalin for 48 h. Fixed stomachs were transferred to 70% ethanol for long-term storage. Prey items were identified to the lowest possible taxon (LPT), enumerated, and weighed (mg). Stomachs containing only parasites were considered "empty" as defined by Manooch et al. (1984); however, these stomachs were still used to assess overall vacuity. Percent by weight (%*W*), percent by number (%*N*), and percent frequency of occurrence (%*O*) were calculated for each LPT. Because some metrics can overinflate the importance of small, numerous prey items (Hyslop 1980), a percent index of relative importance (%IRI) was also calculated (Cortes al. 1997):

$$\%$$
IRI<sub>a</sub> =  $\frac{100 \times IRI}{\sum_{a=1}^{n} IRI}$ 

where  $IRI_a = (\%N + \%W) \times \%O$  and *a* represents samples from 1 to *n*. Gastric endoparasites were identified and the overall prevalence of infection was quantified.

Sample-size sufficiency across ontogeny was assessed with a cumulative prey curve (Ferry and Caillet 1996). The curve was created in PRIMER version 6 and determined the maximum number of expected prey taxa ( $S_{max}$ ) and the actual number of prey taxa across samples ( $S_{obs}$ ) based on the LPT level of identification. To remove the effect of sampling chronology on curve smoothness, the order was randomized across 999 permutations. Sample-size sufficiency in explaining dietary breadth (i.e., asymptotic characteristics) was examined by visual inspection.

Ontogenetic and sex-associated trends in diet composition were assessed using multivariate techniques. We conducted these analyses on prey weights, which were standardized to fish size by dividing the weight of each individual prey item by the individual body weight of the fish (Ajemian and Powers 2012). Standardized prey group weights and numbers were imported into PRIMER, fourth-root transformed, and used to develop a Bray-Curtis similarity matrix. A two-way, crossed permutational, multivariate analysis of variance (PERMANOVA) was used to evaluate differences in diet composition among the various size-classes and sexes (Anderson 2001). Individual Dolphinfish were assigned to one of five size-classes based on 25-cm increments. These size-classes were chosen as they may represent potential ontogenetic shifts that occur with different year-classes and sexual maturity (Young 2014). Binning allowed for an adequate number of samples for each class and a comparison with those of Manooch et al. (1984) who used 20-cm bins. All tests were permutated 999 times under a reduced model (Anderson 2001). Significant factors were further analyzed using PERMANOVA pairwise comparisons, and similarity percentage (SIMPER) analysis was employed to examine the prey items most responsible for the separation among factors (Clarke 1993). We accompanied our analysis with a distance-based test for homogeneity of multivariate dispersions (PERMDISP) to determine possible reasons for the rejection of the null hypothesis (no differences in diet composition) by PERMANOVA, as this test can be sensitive to sample dispersion (Anderson 2006).

#### RESULTS

Of the 357 stomachs collected 281 contained identifiable prey items. For each size-class the following sample sizes were analyzed: 25–49 cm (n = 44), 50–74 cm (n = 199), 75–99 cm (n = 70), 100–124 (n = 23), and 125–149 (n = 21). The cumulative prey curve approached an asymptote, which suggested that the sample size was sufficient to describe the diet (maximum number of unique taxa, or  $S_{obs} = 33$ ,  $S_{max} = 33.74$ ; Figure 1). Fish size ranged from 27.6 to 148.5 cm TL (Figure 2). Sex distribution was 205 females (57%), 134 males (38%), and 18 (5%) undetermined. No prev items were found in 77 individuals (22%). The majority of empty stomachs were from fish of unidentifiable sex (72%), followed by female fish (20%)and males (18%). The smallest size-class (25-49 cm) had the greatest percentage of empty stomachs (32%), followed by the largest size-class (125-149 cm, 29%), the 50-74-cm size-class (22%), the 75–99-cm size-class (16%), and the 100–124-cm size-class (13%; Figure 2). Four major taxonomic classes of prey were identified: Cephalopoda, Malacostraca, Osteichthyes, and Testudines (Table 1). Overall, sargassum occurred in 26% of all stomachs. Sargassum was most common item in the largest size-class (125-149 cm; 43%), followed by the 100-124-cm (30%), 50-74-cm (25%), 75-99-cm (24%), and 25-49-cm (20%) size-classes. Sargassum was more common in males (31%), than in females (24%).



FIGURE 1. Cumulative prey curve plotting mean ( $\pm$ SE) of unique prey items and number of specimens examined for Dolphinfish (n = 357) collected in the western Gulf of Mexico ( $S_{obs} = 33, S_{max} = 33.74$ ).

Pooled data for all size-classes of Dolphinfish indicated a diet dominated by bony fishes (Osteichthyes), regardless of the metric used. By number, Osteichthyes was the most important prey class (70.57%N; Table 1), followed by Malacostraca (24.85%N), Cephalopoda (3.45%N), and Testudines (1.28%N). Within Osteichthyes, the most dominant fish families by number were Carangidae (9.53%N), Tetraodontidae (4.88%N), Monocanthidae (4.88%N), and Balistidae (2.63%N). By weight, Osteichthyes was also the most important prey class (80.80% W), followed by Testudines (2.05% W),Malacostraca (1.15% W),and Cephalopoda (0.47% W; Table 1). Osteichthyan families contributing the most by weight included Carangidae (17.17% W), Monocanthidae (7.42%W), Tetraodontidae (5.43%W), and Balistidae (2.98%W). Using the compound metric %IRI, Osteichthyes was the most important prey class (96.51%IRI), followed by Malacostraca (3.15%IRI), Cephalopoda (0.31%IRI), and Testudines (0.01%IRI; Table 1). The Osteichthyes family with the highest %IRI was Carangidae (4.86%IRI), followed by Monocanthidae (1.26%IRI), Tetraodontidae (1.04%IRI), and Balistidae (0.49%IRI; Figure 3).

When diets were analyzed by size-class (25-cm bins), an ontogenetic shift was observed. By number, Malacostraca was

the most important prey item (68.17%N) for the smallest sizeclass (25-49 cm). However, by weight, this prey group was less important (7.24% W: Table 2). Malacostraca was dominated by small megalopae, classified as Brachyura (39.46%N), which were often consumed in large quantities (Table 2), as well as other small, unidentifiable crustaceans (Decapoda, 26.01%N). For all but the smallest size-class, the diet was over 50% Osteichthyes by number, with a much lower percentage of malacostracans (Table 2). While Osteichthyes was the primary prey category in most size-classes (except 25-49 cm) for all metrics, we observed variability in the importance of different fish families among size-classes. Based on %IRI, Carangidae was the most important identifiable fish family for the smallest sizeclasses (25-49 cm, 50-74 cm, and 75-99 cm); however, for the 100-124-cm size-class, Balistidae was the most important (3.93%IRI) and for 125-149-cm size-class, Tetraodontidae (17.10%IRI) and Monocanthidae (12.57%IRI) were most important (Figure 3). One incident of cannibalism was observed in the 50–74-cm size-class (0.02%IRI). Consumption of nine juvenile Kemp's Ridley sea turtles Lepidochelys kempii was found in a single individual measuring 148 cm TL (3.92%IRI). The turtles ranged in size from 35.4 to 47.5 mm in carapace length.



FIGURE 2. Distribution by size-class (25-cm bins) for TLs (cm) of Dolphinfish collected showing empty stomachs and stomachs containing prey; size-classes were 25–49 cm (n = 44), 50–74 cm (n = 199), 75–99 cm (n = 70), 100–124 cm (n = 23), and 125–149 cm (n = 21).

A two-way crossed PERMANOVA on standardized weights of LPT found no significant interaction between the factors sex and size-class (PERMANOVA: Pseudo- $F_{4, 171} = 1.341$ , P =0.059; Table 3). Assessing the two factors individually, sex was not significant (PERMANOVA: Pseudo- $F_{1, 171} = 1.044$ , P =0.383); however, size-class was significant (PERMANOVA: Pseudo- $F_{4, 171} = 3.269, P = 0.001$ ; Table 3). Subsequent pairwise comparisons indicated that all size-classes were significantly different from each other except the two largest, 100-124 cm and 125–149 cm (PERMANOVA: t = 1.2123, P = 0.155; Table 4). The PERMDISP analysis found that these size-class differences were not explained by sample dispersion (PERMDISP: P = 0.479), and all size-classes had similar mean deviations from the centroid. The SIMPER analysis showed the main contributors to the dissimilarities between the smallest (25-49 cm) and the three largest size-classes (75–99, 100–124, and 125-149 cm) to be a greater abundance of brachyurans and carangids in the diet of the smaller fish (Table 4). The presence of squid in diets of the 100-124-cm size-class and tetraodontids in the 125-149-cm size-class also contributed to these dissimilarities (Table 4).

The overall gastric parasite infection rate was 68%. Prevalence varied by size-class, and there was a general trend of increased parasite burden with increased TL (Figure 4). However, percent occurrence was highest for the 100-cm bin. Two endogastric parasites were identified: the hemurid digenean, *Dinurus* sp., (55.16% rate of infection) and the nematode, *Hysterothylacium pelagicum*, (16.37% rate of infection). There were two occurrences of a digenean parasite that could not be identified beyond subclass.

#### DISCUSSION

Similar to other locations, Dolphinfish in the western Gulf of Mexico are opportunistic predators with a diet dominated by fishes. The diet of this very mobile fish includes a wide array of fish families from the pelagic realm, which is congruent with its habitat use patterns as Dolphinfish are known to spend 60% of their time in the surface layer of the ocean (Merten et al. 2014b). We found ontogenetic shifts in prey for both fish and nonfish prey items; however, unlike some other locations, there were no sex-related differences observed in the diet for this region. Overall, though there were notable dietary changes throughout ontogeny, Dolphinfish in the Gulf of Mexico appear highly dependent on the pelagic environment for food resources.

An ontogenetic shift was observed around sexual maturity (50–74 cm) for both males and females, where diets transitioned from brachyrans, carangids, and balistids to tetraodontids,

#### BREWTON ET AL.

TABLE 1. Overall composition of Dolphinfish (n = 357) diet by prey class and lowest possible taxon showing percent frequency of occurrence (%O), percent by number (%N), percent by weight (%W), and index of relative importance (%IRI). Calculations for %IRI were performed without innumerable contents (i.e., sargassum, marine trash, and unidentified contents). Values in bold text represent totals for prey class. NI = not identifiable beyond major taxon.

Cephalopoda         Teuthida         Teuthida         Teuthida         2.66         3.20         0.47           Malacostraca         14.97         23.00         1.15         14.97         23.00         1.15           Amphipoda, NI         Amphipoda         1.12         1.18         0.04         0.41           Brachyura         Brachyura         Brachyura         7.13         11.67         0.41           Caridea         Caridea         0.14         0.70         0.01         0.04           Decapoda, NI         Decapoda         1.54         4.17         0.04           Hippolytidae         Latreutes parvulus         0.28         0.14         0.01           Isopoda, NI         Isopoda         2.10         1.46         0.04           Penaeidae         Farfantepenaeus sp.         0.28         0.14         0.24         0.07           Portunidae         Callinectus sapidus         0.14         0.21         0.07         0.07           Portunidae         0.84         1.39         0.17         0.06         0.98         1.74         0.06           Osteichthyes         Istention of the section         Istention of the section         0.98         1.74         0.06 <th>%IRI</th>	%IRI
Malacostraca       14.97       23.00       1.15         Amphipoda, NI       Amphipoda       1.12       1.18       0.04         Brachyura       Brachyura       Brachyura       7.13       11.67       0.41         Caridea       Caridea       0.14       0.70       0.01       0.01         Decapoda, NI       Decapoda       1.54       4.17       0.04         Hippolytidae       Latreutes parvulus       0.28       0.14       <0.01         Isopoda, NI       Isopoda       2.10       1.46       0.04         Penaeidae       Farfantepenaeus sp.       0.28       0.14       0.24       0.07         Portunidae       Callinectus sapidus       0.14       0.21       0.07       0.07         Portunidae       Callinectus sapidus       0.42       0.21       0.07       0.07         Stomatopoda, NI       Stomatopoda       0.98       1.74       0.06       0.98       0.74	0.31
Amphipoda, NI       Amphipoda       1.12       1.18       0.04         Brachyura       Brachyura       7.13       11.67       0.41         Caridea       Caridea       0.14       0.70       0.01       0.01         Decapoda, NI       Decapoda       1.54       4.17       0.04         Hippolytidae       Latreutes parvulus       0.28       0.14       <0.01         Isopoda, NI       Isopoda       2.10       1.46       0.04         Penaeidae       Farfantepenaeus sp.       0.28       0.14       0.24         Portunidae       Callinectus sapidus       0.14       0.21       0.07         Portunidae       Callinectus sapidus       0.14       0.21       0.07         Stomatopoda, NI       Stomatopoda       0.98       1.74       0.06	3.15
Brachyura       Brachyura       7.13       11.67       0.41         Caridea       Caridea       0.14       0.70       0.01         Decapoda, NI       Decapoda       1.54       4.17       0.04         Hippolytidae       Latreutes parvulus       0.28       0.14       <0.01	0.04
Caridea       Caridea       0.14       0.70       0.01       0.01         Decapoda, NI       Decapoda       1.54       4.17       0.04         Hippolytidae       Latreutes parvulus       0.28       0.14       <0.01	2.71
Decapoda, NI         Decapoda         1.54         4.17         0.04           Hippolytidae         Latreutes parvulus         0.28         0.14         <0.01	< 0.01
Hippolytidae       Latreutes parvulus       0.28       0.14       <0.01         Isopoda, NI       Isopoda       2.10       1.46       0.04         Penaeidae       Farfantepenaeus sp.       0.28       0.14       0.24         Portunidae       Callinectus sapidus       0.14       0.21       0.07         Portunidae       0.84       1.39       0.17         Portunus sayi       0.42       0.21       0.07         Stomatopoda, NI       Stomatopoda       0.98       1.74       0.06	0.20
Isopoda, NI       Isopoda       2.10       1.46       0.04         Penaeidae       Farfantepenaeus sp.       0.28       0.14       0.24       4         Portunidae       Callinectus sapidus       0.14       0.21       0.07       4         Portunidae       0.84       1.39       0.17       4       4         Portunidae       0.84       1.39       0.17       4         Portunus sayi       0.42       0.21       0.07       4         Osteichthyes       67.27       65.25       80.01       9	< 0.01
Penaeidae         Farfantepenaeus sp.         0.28         0.14         0.24         90.28           Portunidae         Callinectus sapidus         0.14         0.21         0.07         90.28           Portunidae         O.84         1.39         0.17         90.21         0.07         90.21           Portunidae         O.84         1.39         0.17         90.17         90.17         90.17           Stomatopoda, NI         Stomatopoda         0.98         1.74         0.06         90.17           Osteichthyes         67.27         65.25         80.01         90.17	0.10
Portunidae         Callinectus sapidus         0.14         0.21         0.07         0.7           Portunidae         0.84         1.39         0.17         0.06         0.98         1.74         0.06         0.06         0.98         1.74         0.06         0.06         0.06         0.01         0.05         0.01         0.01         0.01         0.01         0.01         0.01         0.01 <t< td=""><td>&lt; 0.01</td></t<>	< 0.01
Portunidae         0.84         1.39         0.17           Portunus sayi         0.42         0.21         0.07         0.07           Stomatopoda, NI         Stomatopoda         0.98         1.74         0.06           Osteichthyes         67.27         65.25         80.01         9	< 0.01
Portunus sayi         0.42         0.21         0.07           Stomatopoda, NI         Stomatopoda         0.98         1.74         0.06           Osteichthyes         67.27         65.25         80.01         9	0.04
Stomatopoda, NI         Stomatopoda         0.98         1.74         0.06           Osteichthyes         67.27         65.25         80.01         9	< 0.01
Osteichthyes 67.27 65.25 80.01	0.06
	6.49
Actinopterygii, NI Actinopterygii 37.34 38.99 33.59	\$8.60
Balistidae Balistidae 2.94 2.29 2.75	0.49
Canthidermis sufflamen 0.14 0.139 0.23	< 0.01
Belonidae Belonidae 0.14 0.07 0.68	(0.01
Carangidae Carangidae 7.41 8.83 10.61	4.74
<i>Caranx crysos</i> 0.14 0.07 0.30	:0.01
<i>Caranx</i> sp. 0.28 0.278 1.36	0.02
Chloroscombrus chrysurus 0.14 0.069 0.01	< 0.01
Decapterus punctatus 0.14 0.14 1.30	0.01
<i>Decapterus</i> sp. 0.28 0.21 0.72	0.01
Hemicaranx amblyrhynchus 0.98 1.53 0.36	0.06
Oligoplites saurus 0.14 0.14 0.14	0.01
Selar crumenophthalmus 0.42 0.21 0.75	0.01
Seriola rivoliana 0.14 0.07 1.62	0.01
Clinidae Clinidae 0.14 0.14 0.10	0.01
Clupeidae Clupeidae 0.56 0.28 2.15	0.05
Jenkinsia lamprotaenia 0.14 0.07 1.07	0.01
Coryphaenidae Coryphaena hippurus 0.14 0.07 1.28	0.01
Diodontidae Diodon holocanthus 0.70 0.49 0.01	< 0.01
Diodon hystrix 0.28 0.14 0.05	< 0.01
<i>Diodon</i> sp. 0.14 0.07 0.31	0.01
Exocoetidae Exocoetidae 1.26 0.70 1.92	0.11
Haemulidae Orthopristis chrysoptera 0.14 0.07 0.51	< 0.01
Hemiramphidae Hemiramphidae 0.14 0.07 0.57	0.01
Hemiramphus brasiliensis 0.28 0.14 0.88	0.01
Istiophoridae Istiophorus platypterus 0.14 0.07 <0.01	0.01
Kyphosidae Kyphosus sectatrix 0.14 0.07 <0.01	< 0.01
Monacanthidae Aluterus scriptus 0.14 0.14 0.43	0.01
<i>Aluterus</i> sp. 0.14 0.07 <0.01	0.01
Cantherhines sp. 4.48 3.06 4.95	1.19
Monacanthidae 0.84 0.49 2.03	0.07
Oxymonacanthus longirostris 0.14 0.07 <0.01	(0.01
Stephanolepis hispidus 0.14 0.07 0.01	<0.01
Pomatomidae Pomatomus saltatrix 0.14 0.14 0.26	<0.01

Prey class	Major taxon	Lowest possible taxon	%O	%N	%W	%IRI
	Scaridae	Scaridae	0.14	0.28	0.04	< 0.01
	Scombridae	Scombridae	0.56	0.35	0.18	0.01
	Sparidae	Lagodon rhomboides	0.28	0.21	1.96	0.02
	1	Sparidae	0.14	0.07	0.58	< 0.01
	Syngnathidae	<i>Hippocampus</i> sp.	0.14	0.07	0.02	< 0.01
	Tetraodontidae	Lagocephalus laevigatus	0.14	0.28	0.35	< 0.01
		Sphoeroides sp.	0.28	0.83	0.09	0.01
		Tetraodontidae	3.78	3.27	4.98	1.03
	Tetraodontiformes, NI	Tetraodontiformes	0.56	0.28	0.05	0.01
	Trichiuridae	Trichiurus lepturus	0.14	0.07	0.15	< 0.01
	Triglidae	Triglidae	0.28	0.14	0.66	0.01
Testudines	Cheloniidae	5	0.28	1.18	2.05	0.01
		Cheloniidae	0.14	0.56	0.01	< 0.01
		Lepidochelvs kempii	0.14	0.63	2.04	0.01
Phaeophyceae	Sargassaceae	Sargassum sp.	12.87	6.39		
Marine trash	8	0	1.96	0.97		
Unidentified					15.50	

TABLE 1. Continued.

monocanthids, and squid (teuthida). This dietary shift may be a result of ontogenetic changes in habitat use. Mature Dolphinfish make deep dives as night, which may provide increased opportunities for larger fish to find more diverse prey resources (Merten et al. 2014b). It is also possible that squid are being consumed as they make diel vertical migrations towards the surface at night (Passarella and Hopkins 1991), which may make them more available to epipelagic fishes.

In our study, the prevalence of the two of the most commonly observed fish families, Carangidae and Monocanthidae, in the diets had divergent trends with Dolphinfish TL. Specifically, carangids were most common in diets of the smallest size-class, while monocanthids increased in importance with the larger size-classes. This dietary shift may be the result of changing foraging strategies with age (Nunes et al. 2015) or simply gape limitation. Ontogenetic shifts in Dolphinfish diets have been observed in other regions; however, specific trends in feeding have varied by location (Manooch 1984; Tripp-Valdez 2015). Our study suggests that Dolphinfish feeding ecology and habitat use are highly dynamic and vary ontogenetically throughout the species' range.

We also observed dietary shifts when examining nonfish prey items. Brachyuran megalopae were numerically dominant in smaller size-classes, where large quantities of these small prey items (up to 53 individuals) were often consumed. Other studies from the Gulf of California have shown Dolphinfish consume large quantities of megalopae, presumably due to availability and ease of capture (Tripp-Valdez et al. 2010), both of which may be important for these faster-growing, small size-classes. Conversely, in larger size-classes of Dolphinfish, swimming crabs (Portunidae) were numerically dominant for the prey class Malacostraca. Dolphinfish may require more skill to capture these more elusive prey than drifting planktonic megalopae. These findings emphasize that ontogenetic change in feeding ecology may also be driven by the increased feeding agility of fish from larger size-classes, as reported by Nunes et al. (2015).

This is the first documented report of the presence of endangered Kemp's Ridley sea turtles in the Dolphinfish diet. Although sea turtles were found in only a single Dolphinfish in the present study, reports from regional online fishing boards have also documented sea turtle consumption in the Gulf of Mexico (e.g., see http://www.thefishingwire.com/ features/224956). The proximity of Texas to the only known natural Kemp's Ridley sea turtle nesting site (Rancho Nuevo, Tamaulipas, Mexico) and local statewide head start (Caillouet et al. 2015) and other hatchling release programs may cause Dolphinfish to encounter relatively higher densities of hatchlings as they disperse from nests along Gulf of Mexico shorelines. Indeed, turtles observed in this study were at hatchling size (Marquez 1994). Similarly, sea turtle hatchling predation has also been documented in the Atlantic Ocean in the diet of Atlantic Sharpnose Sharks Rhizoprionodon terraenovae (Delorenzo et al. 2015). These young sea turtles are also known to associate with pelagic sargassum in this region (Witherington et al. 2012), which may aggregate them in preferred Dolphinfish habitat (see below). Additionally, anecdotal reports from commercial fishers in Oaxaca, Mexico, describe sea turtle predation by Dolphinfish as a fairly



FIGURE 3. Percent by weight (%*W*) for prey families or lowest possible taxon (LPT) observed in Dolphinfish diets by size-class (cm TL); size-classes were 25–49 cm (n = 44), 50–74 cm (n = 199), 75–99 cm (n = 70), 100–124 cm (n = 23), and 125–149 cm (n = 21). LPTs contributing <5% were combined by taxonomic class: Combined Fish (Clinidae, Istiophoridae, Kyphosidae, Pomatomidae, Scaridae, Scombridae, Sygnathidae, Tetradontiformes, and Trichiuridae) and Combined Crustaceans (Amphipoda, Caridea, Decapoda, Hippoltidae, Isopoda, Portunidae, and Stomatopoda).

common occurrence in the eastern Pacific Ocean as well (F. Galguera, Puerto Angel, personal communication). These convergent findings suggest that the Dolphinfish is an important chelonian predator and may feed on more diverse resources than previously considered.

The nondiscriminatory feeding habits of Dolphinfish are evidenced by the fairly common occurrence of nonfood items in the stomach contents. Such items included pieces of rope, plastic ribbons, hard plastic bits, and a badminton shuttlecock. This nondiscriminatory feeding style has also been described in many other locales (Gibbs and Collette 1959; Rose and Hassler 1974; Manooch et al. 1984; Rudershausen et al. 2010; Varghese et al. 2013). The most common nonfood item, sargassum, was observed in 26% of stomachs analyzed, and the presence of sargassum in stomach contents was found to increase with Dolphinfish TL. These findings suggest that Dolphinfish increase their use of sargassum habitat with age. However, all fish size-classes had prey items associated with sargassum habitats in the Gulf of Mexico (Wells and Rooker 2004b; Hoffmayer et al. 2005; Taylor et al. 2015). Therefore, it is likely that sargassum is ingested while feeding on fish and other organisms using this floating structure as habitat, and this may be more conspicuous in larger individuals capable of

ingesting larger pieces of this macroalgae. It is also possible that some nutrition may be gained from the ingestion of plant material as suggested by Bethea at al. (2007) in their analysis of Bonnethead Sphyrna tiburo diets. Therefore, future diet studies may want to further investigate the nutritional contribution of sargassum to Dolphinfish. Sargassum forms large lines and mats offshore and supports a diverse community of transient and juvenile fishes and invertebrates that comprise an important prey base for pelagic fish predators (SAFMC 2002; Wells and Rooker 2004b; Hoffmayer et al. 2005; Taylor et al. 2015). In previous studies the presence of sargassum was observed to be a common occurrence in Dolphinfish stomach contents, ranging from 28% (Rose and Hassler 1974) to 48% (Manooch 1984), and throughout size-classes (Ruderhausen et al. 2010). Our findings and previous work documenting consumption of floating nonfood items and sargassum confirm Dolphinfish reliance on dynamic pelagic habitats. As per Manooch et al. (1984), this feeding behavior may make this economically important species vulnerable to anthropogenic impacts, such as oil spills, along the ocean surface.

We observed no differences in prey consumed between sexes; however, in the Gulf of Mexico, slower growth rates and younger sexual maturity have been estimated for females

6
<u> </u>
0
$\overline{\sim}$
$\geq$
<u>د</u>
<b>_</b>
$\sim$
<u> </u>
-
-
•••
_
—
<u> </u>
E,
_
1
E C
Ч
5
$\cup$
$\mathbf{S}$
Ξ.
5.
-
5
r
$\cup$
$\sim$
5
·=
~~
SL
/ers
ivers
nivers
Jnivers
Univers
[ Univers
<b>A</b> Univers
M Univers
&M Univers
&M Univers
A&M Univers
: A&M Univers
ts A&M Univers
as A&M Univers
xas A&M Univers
exas A&M Univers
Texas A&M Univers
[Texas A&M Univers
/ [Texas A&M Univers
y [Texas A&M Univers
by [Texas A&M Univers
d by [Texas A&M Univers
ed by [Texas A&M Univers
led by [Texas A&M Univers
ded by [Texas A&M Univers
aded by [Texas A&M Univers
oaded by [Texas A&M Univers
nloaded by [Texas A&M Univers
nloaded by [Texas A&M Univers
wnloaded by [Texas A&M Univers
ownloaded by [Texas A&M Univers
Downloaded by [Texas A&M Univers

TABLE 2. Overall composition by size-class (25-cm bins) of Dolphinfish diet by prey class and lowest possible taxon showing percent by number (%/N), percent by weight (%/W), and index of relative importance (%IRI); sample sizes for bins were 25-49 cm (25; n = 44), 50–74 cm (50; n = 199), 75–99 cm (75; n = 70), 100–124 (100; n = 23), and 125–149 (125; n = 21). Values in bold text represent totals for major prey class (see Table 1).

		Percent	by numbe	er (%N)			Percent	by weigh	it (%W)		Inde	κ of relati	ve impor	tance (%]	RI)
Taxonomic group	25	50	75	100	125	25	50	75	100	125	25	50	75	100	125
Cephalopoda, Teuthida		1.21	5.31	18.75	2.63		0.79	0.05	1.27	0.10		0.06	0.43	5.93	0.37
Malacostraca	68.17	16.89	13.47	7.14	10.53	7.24	1.83	0.16	0.10	2.13	18.18	1.49	1.77	0.49	3.40
Amphipoda		2.28					0.11					0.10			
Brachyura	39.46	7.92	7.76	0.89	0.88	4.34	0.54	0.13	<0.01	0.65	12.28	1.22	1.65	0.04	0.10
Caridea			4.08					0.03					0.07		
Decapoda	26.01	0.13	0.41			1.13	0.05	<0.01			5.69	<0.01	0.01		
Hippolytidae		0.13			0.88		<0.01			<0.01		<0.01			0.06
Isopoda	1.35	1.74		3.57	0.88	0.13	0.10		0.03	<0.01	0.10	0.10		0.18	0.06
Penaeidae	0.45	0.13				1.47	0.60				0.07	<0.01			
Portunidae	0.45	2.01	0.41		7.89	0.01	0.27	<0.01		1.48	0.02	0.04	0.01		3.18
Stomatopoda	0.45	2.55	0.82	2.68		0.15	0.15	0.01	0.07		0.02	0.03	0.03	0.27	
Osteichthyes	27.80	74.45	71.41	66.91	63.17	78.16	83.45	86.76	70.28	70.25	64.86	85.93	78.31	58.47	69.93
Actinopterygii	16.14	49.26	33.06	36.61	31.58	47.15	44.13	31.95	29.24	15.23	57.23	78.79	59.46	46.74	38.23
Balistidae	0.45	1.61	4.49	8.04	1.75	2.90	2.99	4.57	7.43	1.13	0.46	0.47	0.90	3.93	0.39
Belonidae					0.88					4.58					0.38
Carangidae	6.73	12.35	22.04	2.68	1.75	13.32	12.97	32.27	7.29	0.14	5.78	5.58	16.36	1.03	0.26
Clinidae				1.79					0.76					0.13	
Clupeidae		0.40	0.41	0.89			4.06	2.10	8.12			0.07	0.04	0.47	
Coryphaenidae		0.13					3.71					0.02			
Diodontidae		0.13	0.41	2.68	2.63		0.46	0.87	3.38	1.65		<0.01	0.02	0.62	0.58
Exocoetidae	1.79	0.40	0.82		0.88	13.50	1.33	1.66		4.15	1.11	0.04	0.08		0.35
Haemulidae		0.13					1.49					0.01			
Hemiramphidae		0.13	0.38	0.83			1.30	1.61	3.23			0.01	0.03	0.21	
Istiophoridae		0.13					0.01					<0.01			
Kyphosidae		0.13					0.01					<0.01			
Monacanthidae	2.24	3.76	4.08	4.46	7.02	1.13	3.95	3.23	3.98	23.41	0.26	0.50	0.48	2.14	12.57
Pomatomidae		0.27					0.74					0.01			

#### DOLPHINFISH DIET IN THE GULF OF MEXICO

		Percent t	y numbe	er (%N)			Percent b	y weigh	t (%W)		Index	κ of relati	ve impor	tance (%	(IRI)
Taxonomic group	25	50	75	100	125	25	50	75	100	125	25	50	75	100	125
Scaridae		0.54					0.13					<0.01			
Scombridae	0.45	0.40	0.41			0.18	0.10	0.41			0.02	0.01	0.01		
Sparidae		0.27	0.82				3.75	3.52				0.02	0.15		
Syngnathidae		0.13					0.05					<0.01			
Tetraodontidae		3.76	3.67	8.93	15.79		1.78	2.67	6.87	19.88		0.40	0.73	3.20	17.10
Tetraodontiformes		0.27	0.41		0.88		0.04	0.08		0.09		<0.01	0.01		0.07
Trichiuridae		0.13					0.45					<0.01			
Triglidae		0.13	0.41				0.03	1.83				<0.01	0.04		
Testudines, Cheloniidae					14.91					13.77					3.92
Sargassum	4.04	7.38	9.80	7.14	8.77										
Unidentified	8.55	11.65	15.33	13.33	11.11	14.60	13.94	13.04	28.34	13.76	16.95	12.51	19.47	35.11	22.39

TABLE 2. Continued.

Source	df	SS	MS	Pseudo-F	P (permutation)	Unique permutations
Size-class	4	51,446	12,861	3.269	0.001	999
Sex	1	4,106.2	4,106.2	1.044	0.383	997
Size-class × Sex	4	21,101	5,275.2	1.341	0.059	995
Residual	162	$6.37 \times 10^{5}$	3,934.8			
Total	171	$7.21 \times 10^{5}$				

TABLE 3. Results from a two-way crossed PERMANOVA of Dolphinfish dietary composition among size-classes (25-cm bins) and sex; SS = sum of squares, MS = mean sum of squares.

(Young 2014). As diet composition was similar between sexes, differing growth rates suggest that females in this region are devoting energy to processes other than somatic growth. For example, female Dolphinfish in the Gulf of Mexico may be dedicating more energy towards reproduction than males. In the Pacific Ocean, females can reach maturity at smaller sizes (Alejo-Plata et al. 2011), and in that region similar feeding ecology between sexes has also been observed through stomach content (Tripp-Valdez et al. 2010; Torres-Rojas et al. 2014) and stable isotope analysis (Tripp-Valdez et al. 2015). As such, despite varying growth rates and potential differences in energetic requirements, males and females still appear to occupy similar ecological feeding niches across a large portion of the species' range.

We found Dolphinfish to have a high gastric parasite burden that increased with fish size, congruent with the findings of Burnett-Herkes (1974) and Manooch (1984). The endoparasites present in this research were similar to those found in other locales globally (Carbonell et al. 1999). The most prevalent gastric parasite we observed, Dinurus, has been observed in Dolphinfish from the Mediterranean and Caribbean seas (Carbonell et al. 1999) and is a characteristic parasite of the species (Williams and Bunkley-Williams 2009). The proposed life cycle for *Dinurus* suggests that either benthic gastropods or planktivorous fishes (e.g., clupeids) act as intermediate hosts, of which Dolphinfish are the definitive host (Carbonell et al. 1999). This scenario appears plausible in the Gulf of Mexico given that the size-class with the highest occurrence of clupeid fish in the diet (i.e., 100-124 cm) had the greatest prevalence of *Dinurus* infection. The second most commonly observed parasite, Hysterothylacium pelagicum, is also endemic to Dolphinfish (Williams and Bunkley-Williams 2009) and has been previously observed in the Gulf of Mexico, Atlantic and Pacific oceans, and Caribbean Sea (Deardorff and Overstreet 1982; Manooch 1984; Dyer et al. 1997). The intermediate hosts of parasites from the genus Hysterothylacium are not well defined, although the parasites are thought to reach sexual maturity in the digestive tract of bony fishes or other vertebrates (Marcogliese 1995). In this study, only the stomach was examined. Thus, any additional parasites occupying the intestines would not have been accounted for. As H. pelagicum inhabits both the stomach cavity and intestines (Williams and Bunkley-Williams 2009),

the prevalence observed in this study is likely an underestimation. Nonetheless, the overall percentage of gastric parasitism observed (54%) was higher than the 5% found by Manooch et al. (1984) in the same region. This difference could be due to increases in prevalence of the parasite for the region over the last 30 years, or to our larger sample size (122 versus 357 individuals, respectively). Parasite species have the potential to be identified using molecular methods. Unfortunately, since all stomachs in this study were fixed in formalin, which degrades DNA, this approach was not possible. Future studies may consider alternative stomach and intestinal fixation protocols in order to apply molecular methods to further refine these parasite and dietary analyses. A complete parasitic assessment in this region would allow for a better understanding of the health of the Dolphinfish population as well as its trophic position in the offshore food web.

The families of fish that dominated the observed diet in this study (Carangidae, Balistidae, Monocanthidae, and Tetraodontidae) are common to offshore, pelagic habitats (SAFMC 2002; Wells and Rooker 2004b; Taylor et al. 2015). However, these families also have unique morphological characteristics that make them identifiable even in advanced states of decomposition. For example, carangids can be identified by caudal scutes, balistids (and monocanthids) by location and number of dorsal spines, and tetraodontids by their unusual beak (Hoese and Moore 1998). This greater conspicuousness, even as most scales, fin rays, and other morphological characters have been digested, may have potentially inflated the importance of these families in the diet. Depending on sizeclass, anywhere from 15% to 47% of prey fish were not identifiable beyond the superclass Osteichthyes. Similar to parasite studies, continued research into the feeding habits in the region would be greatly enhanced by the inclusion of DNA-based identification (i.e., barcoding), which has proven useful in understanding marine fish diets by successfully identifying prey fish in advanced states of digestion (Valdez-Moreno et al. 2012; Cote et al. 2013).

Dolphinfish are an important fisheries species with a high dependence on pelagic resources throughout ontogeny. The primary components of their diet in the western Gulf of Mexico included fish from the families Balistidae, Monacanthidae, and Carangidae. Some of these prey items are likely fishery species including Gray Triggerfish *Balistes capriscus* and Greater

#### BREWTON ET AL.

TABLE 4. Results from a two-way, crossed similarity percentage (SIMPER) analyses for significant effects by size-class on Dolphinfish dietary data. Average abundances are multiplied by  $1 \times 10^5$ ; sample sizes for total length bins were 25–49 cm (25; n = 44), 50–74 cm (50; n = 199), 75–99 cm (75; n = 70), 100–124 cm (100; n = 23), and 125–149 cm (125; n = 21).

Taxon	Abur	Idance	Average dissimilarity $\pm$ SD	Contribution (%)	Cumulative contribution (%)
		25	versus 50 cm; $t = 1.4908$ , $P = 0.0$	)14	
Average dissimilarity = 87.99	25	50	, , ,		
Carangidae	0.97	0.72	$18.02 \pm 0.80$	20.48	20.48
Brachyura	1.28	0.46	$17.76 \pm 0.88$	20.19	40.66
Balistidae	0.32	0.24	$12.84 \pm 0.56$	14.59	55.25
		25	versus 75 cm; $t = 1.7774$ , $P = 0.0$	002	
Average dissimilarity = 88.91	25	75	, , ,		
Brachyura	1.28	0.41	$20.31 \pm 0.93$	22.84	22.84
Carangidae	0.97	0.63	$18.22 \pm 0.83$	20.49	43.34
Balistidae	0.32	0.19	$10.64 \pm 0.50$	11.96	55.30
		25	versus 100 cm: $t = 1.8932$ , $P = 0.0$	001	
Average dissimilarity $= 97.25$	25	100			
Brachyura	1.28	0.06	$16.79 \pm 0.82$	17.26	17.26
Carangidae	0.97	0.14	$14.04 \pm 0.81$	14.44	31.70
Balistidae	0.32	0.32	$12.70 \pm 0.58$	13.05	44.76
Teuthida	0.00	0.49	$9.51 \pm 0.53$	9.77	54.53
		25	versus 125 cm: $t = 2.2363$ $P = 0.000$	001	
Average dissimilarity = $9535$	25	125	t = 2.2505, t = 0.5	001	
Brachvara	1.28	0.08	$15.27 \pm 0.86$	16.02	16.02
Carangidae	0.97	0.17	$12.27 \pm 0.80$	12.86	28.88
Monacanthidae	0.32	0.52	$9.82 \pm 0.87$	10.30	39.19
Tetraodontidae	0.00	0.61	$9.02 \pm 0.00$ $9.24 \pm 0.74$	9.69	48.87
Portunidae	0.11	0.46	$850 \pm 0.72$	8.91	57 78
Tortunidae	0.11	50.10	$0.50 \pm 0.72$	0.91	51.10
Average dissimilarity = 86.00	50	50 75	versus 75 cm; $t = 1.5172$ , $P = 0.0$	113	
Corongidae	0.72	0.63	$21.52 \pm 0.84$	24 74	24.74
Brochung	0.72	0.05	$21.52 \pm 0.84$	24.74	24.74 42.22
Monacanthidae	0.40	0.41	$15.2 \pm 0.71$	17.47	42.22
Monacantinuae	0.25	0.14	9.33 ± 0.48	10.72	52.94
	50	100	versus 100 cm; $t = 1.922, P = 0.0$	)01	
Average dissimilarity = $92.33$	50 0.72	100	14.28 + 0.72	15 47	15 47
Tauthida	0.72	0.14	$14.28 \pm 0.73$	13.47	13.47
Managanthidag	0.10	0.49	$11.80 \pm 0.58$	12.85	28.32
Delistidae	0.25	0.32	$10.80 \pm 0.39$	11.70	40.01
Banstidae	0.24	0.32	$9.28 \pm 0.46$	10.05	30.07
	50	50	versus 125 cm; $t = 2.2007, P = 0.0$	001	
Average dissimilarity = $89.84$	50	125	14.00 + 0.04	15 (7	15 (7
letraodontidae	0.24	0.61	$14.08 \pm 0.94$	15.67	15.67
Monacanthidae	0.25	0.52	$13.17 \pm 1.00$	14.66	30.33
Carangidae	0.72	0.17	$12.20 \pm 0.81$	13.58	43.91
Portunidae	0.06	0.46	$8.04 \pm 0.64$	8.95	52.85
		75	versus 100 cm; $t = 1.7372$ , $P = 0.0$	001	
Average dissimilarity = $92.12$	75	100			
Carangidae	0.63	0.14	$14.68 \pm 0.74$	15.93	15.93
Teuthida	0.14	0.49	$13.91 \pm 0.65$	15.10	31.03
Balistidae	0.19	0.32	$11.34 \pm 0.52$	12.30	43.34
Tetraodontidae	0.19	0.23	$9.91 \pm 0.51$	10.75	54.09
		75	versus 125 cm; $t = 2.0404, P = 0.0$	001	
Average dissimilarity = $91.53$	75	125			
Tetraodontidae	0.19	0.61	$13.81 \pm 0.92$	15.08	15.08
Carangidae	0.63	0.17	$12.08 \pm 0.82$	13.20	28.28
Monacanthidae	0.14	0.52	$11.73 \pm 0.88$	12.81	41.09
Portunidae	0.03	0.46	$9.92 \pm 0.73$	10.84	51.93



FIGURE 4. Percent occurrence (%*O*) by size-class (cm) of gastric parasites (families Raphidascarididae and Hemiuiridae) from Dolphinfish collected in the western Gulf of Mexico; size-classes were 25–49 cm (n = 25), 50–74 cm (n = 50), 75–99 cm (n = 75), 100–124 cm (n = 100), and 125–149 cm (n = 125).

Amberjack Seriola dumerili. These species are known to settle out of the sargassum community and onto reefs in the Gulf of Mexico (Wells and Rooker 2004a; Simmons and Szedlmayer 2011; Ajemian et al. 2015), thus demonstrating the potential for pelagic-benthic habitat coupling via Dolphinfish predation. Though only one stomach was found to contain Kemp's Ridley sea turtles, this does suggest that Dolphinfish are potential chelonian predators, which must be considered in sea turtle restoration and conservation efforts. Given the highly mobile nature and the estimated age of fish examined in this study, the timing of collection suggests potential exposure to waterborne contaminants from the Deepwater Horizon oil spill occurs during critical life stages (Rooker et al. 2013; Young 2014). As such, it is important to understand the feeding ecology specific to this region as the ecosystem moves towards recovery. Overall, the diet of Dolphinfish reveals interactions with other managed and protected pelagic species, and these data should help managers face the challenges to sustainable management of these highly migratory pelagic species in the western Gulf of Mexico.

#### ACKNOWLEDGMENTS

This research was made possible in part by a grant from the Corpus Christi Rotary Club's Harvey Weil Sportsman Conservationist Award and a grant from The Gulf of Mexico Research Initiative. Data are publicly available through the Gulf of Mexico Research Initiative Information and Data Cooperative (GRIIDC). We thank C. Downey, M. Sluis, J. Pesik, and A. Ferguson for laboratory assistance and M. Andres for the parasite expertise. The stomach samples in this study were collected through a cooperative agreement between the recreational fishing community of Port Aransas, Texas, and the Harte Research Institute for Gulf of Mexico Studies. Given the wide habitat range of pelagic species such as Dolphinfish, this work would not have been possible without the cooperation of these citizen scientists.

#### REFERENCES

- Alejo-Plata, C., P. Díaz-Jaimes, I. H. and Salgado-Ugarte. 2011. Sex ratios, size at sexual maturity, and spawning seasonality of Dolphinfish (*Coryphaena hippurus*) captured in the Gulf of Tehuantepec, Mexico. Fisheries Research 110:207–216.
- Ajemian, M. J., and S. P. Powers. 2012. Habitat-specific feeding by Cownose Rays (*Rhinoptera bonasus*) of the northern Gulf of Mexico. Environmental Biology of Fishes 95:79–97.
- Ajemian, M. J., J. J. Wetz, B. Shipley-Lozano, J. D. Shively, and G. W. Stunz. 2015. An analysis of artificial reef fish community structure along the northwestern Gulf of Mexico Shelf: potential impacts of "Rigs-to-Reefs" programs. PLoS (Public Library of Science) ONE [online serial] 10(5): e0126354.

- Anderson, M. J. 2001. A new method for non-parametric multivariate analysis of variance. Austral Ecology 26:32–46.
- Anderson, M. J. 2006. Distance-based tests for homogeneity of multivariate dispersions. Biometrics 62:245–253.
- Bannister, J. V. 1976. The length–weight relationship, condition factor and gut contents of the Dolphinfish *Coryphaena hippurus* (L.) in the Mediterranean. Journal of Fish Biology 9:335–338.
- Bascompte, J., C. J. Melián, and E. Sala. 2005. Interaction strength combinations and the overfishing of a marine food web. Proceedings of the National Academy of Sciences of the USA 102:5443–5447.
- Baum, J. K., and B. Worm. 2009. Cascading top-down effects of changing oceanic predator abundances. Journal of Animal Ecology 78:699–714.
- Beardsley, G. L. 1967. Age, growth, and reproduction of Dolphin Coryphaena hippurus in the Straits of Florida. Copeia 1967:441–451.
- Benetti, D. D., R. W. Brill, and S. A. Kraul. 1995. The standard metabolic rate of Dolphin Fish. Journal of Fish Biology 46:987–996.
- Bethea, D. M., L. Hale, J. K. Carlson, E. Cortés, C. A. Manire, and J. Gelsleichter. 2007. Geographic and ontogenetic variation in the diet and daily ration of the Bonnethead shark, *Sphyrna tiburo*, from the eastern Gulf of Mexico. Marine Biology 152:1009–1020.
- Burnett-Herkes, J. N. 1974. Parasites of the gills and buccal cavity of the dolphin, *Coryphaena hippurus*, from the Straits of Florida. Transactions of the American Fisheries Society 103:101–106.
- Caillouet, C. W., D. J. Shaver, and A. M. Landry. 2015. Kemp's ridley sea turtle (*Lepidochlys kempii*) head-start and reintroduction to Padre Island National Seashore, Texas. Herpetological Conservation and Biology 10:309–377.
- Carbonell, E., E. Massutí, J. J. Castro, and R. M. Garcka. 1999. Parasitism of dolphinfishes, *Coryphaena hippurus* and *Coryphaena equiselis*, in the western Mediterranean (Balearic Islands) and central-eastern Atlantic (Canary Islands). Scientia Marina 63:343–354.
- Castriota, L., C. Pipitone, S. Campagnuolo, M. Romanelli, A. Potoschi, and F. Andaloro. 2007. Diet of *Coryphaena hippurus* (Coryphaenidae) associated with FADs in the Ionian and southern Tyrrhenian seas. Cybium 31:435–441.
- Clarke, K. R. 1993. Non-parametric multivariate analyses of changes in community structure. Australian Journal of Ecology 18:117–143.
- Cortés, E. 1997. A critical review of methods of studying fish feeding based on analysis of stomach contents: application to elasmobranch fishes. Canadian Journal of Fisheries and Aquatic Science 54:726–738.
- Côté, I. M., S. J. Green, J. A. Morris Jr., J. L. Akins, and D. Steinke. 2013. Diet richness of invasive Indo-Pacific Lionfish revealed by DNA barcoding. Marine Ecology Progress Series 472:249–256.
- Deardorff, T. L., and R. M. Overstreet. 1982. Hysterothylacium pelagicum sp. n. and H. cornutum (Stossich, 1904) (Nematoda: Anisakidae) from marine fishes. Proceedings of the Helminthological Society of Washington 49:246–251.
- Delorenzo, D. M., D. M. Bethea, and J. K. Carlson. 2015. An assessment of the diet and trophic level of Atlantic Sharpnose Shark *Rhizoprionodon terraenovae*. Journal of Fish Biology 86:385–391.
- Dyer, W. G., L. Bunkley-Williams, and E. H. Williams. 1997. Parasites of the Dolphinfish (*Coryphaena hippurus*) in Puerto Rico. Journal of the Helminthological Society of Washington 64:188–194.
- Ferry, L. A., and G. M. Caillet. 1996. Sample size and data analysis: are we characterizing and comparing diet properly? Pages 71–80 in D. MacKinlay and K. Shearer, editors. Gutshop '96: feeding ecology and nutrition in fish symposium proceedings. American Fisheries Society, Physiology Section, Bethesda, Maryland.
- Frank, K. T., B. Petrie, J. S. Choi, and W. C. Leggett. 2005. Trophic cascades in a formerly cod-dominated ecosystem. Science 308:1621–1623.
- Gibbs, R. H., and B. B. Collette. 1959. On the identification, distribution, and biology of the dolphins, *Coryphaena hippurus* and *C. equiselis*. Bulletin of Marine Science 9:117–152.

- Heithaus, M. R., A. Frid, A. J. Wirsing, and B. Worm. 2008. Predicting ecological consequences of marine top predator declines. Trends in Ecology and Evolution 23:202–210.
- Hoese, H. D., and R. H. Moore. 1998. Fishes of the Gulf of Mexico, Texas, Louisiana, and adjacent waters. Texas A&M University Press, College Station.
- Hoffmayer, E. R., J. S. Franks, B. H. Comyns, J. R. Hendon, and R. S. Waller. 2005. Larval and juvenile fishes associated with pelagic *Sargassum* in the northcentral Gulf of Mexico. Proceedings of the Gulf and Caribbean Fisheries Institute 56:259–269.
- Hyslop, E. J. 1980. Stomach contents analysis-a review of methods and their application. Journal of Fish Biology 17:411–429.
- Kitchens, L. L., and J. R. Rooker. 2014. Habitat associations of Dolphinfish larvae in the Gulf of Mexico. Fisheries Oceanography 23:460–471.
- Lasso, J., and L. Zapata. 1999. Fisheries and biology of *Coryphaena hippurus* (Pisces: Coryphaenidae) in the Pacific coast of Colombia and Panama. Scientia Marina 63:387–399.
- Mager, E. M., A. J. Esbaugh, J. D. Stieglitz, R. Hoenig, C. Bodinier, J. P. Incardona, N. L. Schloz, D. D. Benetti, and M. Grosell. 2014. Acute embryonic or juvenile exposure to Deepwater Horizon crude oil impairs the swimming performance of Mahi-mahi (*Coryphaena hippurus*). Environmental Science and Technology 48:7053–7061.
- Manooch, C. S., D. L. Mason, and R. S. Nelson. 1984. Food and gastrointestinal parasites of Dolphin *Coryphaena hippurus* collected along the southeastern and Gulf coasts of the United States. Bulletin of the Japanese Society of Scientific Fisheries 50:1511–1525.
- Marcogliese, D. J. 1995. The role of zooplankton in the transmission of helminth parasites to fish. Reviews in Fish Biology and Fisheries 5:336–371.
- Marcogliese, D. J. 2002. Food webs and the transmission of parasites to marine fish. Parasitology 124:83–99.
- Marquez, M. R. 1994. Synopsis of biological data on the Kemp's ridley turtle, *Lepidochelys kempi* (Garman, 1880). NOAA Technical Memorandum NMFS-SEFSC-343.
- Massutí, E., S. Deudero, P. Sánchez, and B. Morales-Nin. 1998. Diet and feeding of dolphin (*Coryphaena hippurus*) in western Mediterranean waters. Bulletin of Marine Science 63:329–341.
- Massutí, E., and B. Morales-Nin. 1997. Reproductive biology of Dolphin-fish (Coryphaena hippurus L.) off the island of Majorca (western Mediterranean). Fisheries Research 30:57–65.
- Merten, W., R. Appeldoorn, and D. Hammond. 2014a. Movements of Dolphinfish (*Coryphaena hippurus*) along the U.S. East Coast as determined through mark and recapture data. Fisheries Research 151:114–121.
- Merten, W., R. Appeldoorn, R. Rivera, and D. Hammond. 2014b. Diel vertical movements of adult male Dolphinfish (*Coryphaena hippurus*) in the western central Atlantic as determined by use of pop-up satellite archival transmitters. Marine Biology 161:1823–1834.
- Nunes, J. D. A. C., R. H. Freitas, J. A. Reis-Filho, M. Loiola, and C. L. Sampaio. 2015. Feeding behaviour of the common Dolphinfish *Coryphaena hippurus*: older fish use more complex foraging strategies. Journal of the Marine Biological Association of the UK 95:1277–1284.
- Olson, R. J., and F. Galván-Magaña. 2002. Food habits and consumption rates of common Dolphinfish (*Coryphaena hippurus*) in the eastern Pacific Ocean. U.S. National Marine Fisheries Service Fishery Bulletin 100:279–298.
- Oxenford, H. A., and W. Hunte. 1999. Feeding habits of the Dolphinfish (*Coryphaena hippurus*) in the eastern Caribbean. Scientia Marina 63:317–325.
- Palko, B. J., G. L. Beardsley, and W. J. Richards. 1982. Synopsis of the biological data on dolphin-fishes, *Coryphaena hippurus* Linnaeus and *Coryphaena equiselis* Linnaeus. NOAA Technical Report NMFS Circular 443.
- Passarella, K. C., and T. L. Hopkins. 1991. Species composition and food habits of the micronektonic cephalopod assemblage in the eastern Gulf of Mexico. Bulletin of Marine Science 49:638–659.

- Raptopoulou, F. A., and R. H. Lambertsen. 1987. Parasite-associated pathology of the Dolphin fish, *Coryphaena hippurus* L., from Florida waters. Journal of Fish Diseases 10:379–384.
- Rooker, J. R., L. L. Kitchens, M. A. Dance, R. D. Wells, B. Falterman, and M. Cornic. 2013. Spatial, temporal, and habitat-related variation in abundance of pelagic fishes in the Gulf of Mexico: potential implications of the Deepwater Horizon oil spill. PLoS (Public Library of Science) One [online serial] 8(10):e76080.
- Rose, C. D., and W. W. Hassler. 1974. Food habits and sex ratios of dolphin *Coryphaena hippurus* captured in the western Atlantic Ocean off Hatteras, North Carolina. Transactions of the American Fisheries Society 103:94– 100.
- Rudershausen, P. J., J. A. Buckel, J. Edwards, D. P. Gannon, C. M. Butler, and T. W. Averett. 2010. Feeding ecology of Blue Marlins, Dolphinfish, Yellowfin Tuna, and Wahoos from the North Atlantic Ocean and comparisons with other oceans. Transactions of the American Fisheries Society 139:1335–1359.
- SAFMC (South Atlantic Fishery Management Council). 2002. Fishery management plan for pelagic Sargassum habitat of the south Atlantic region. SAFMC, Charleston, South Carolina.
- Schwenke, K. L., and J. A. Buckel. 2008. Age, growth, and reproduction of Dolphinfish (*Coryphaena hippurus*) caught off the coast of North Carolina. U.S. National Marine Fisheries Service Fishery Bulletin 106:82–92.
- Shackell, N. L., K. T. Frank, J. A. Fisher, B. Petrie, and W. C. Leggett. 2010. Decline in top predator body size and changing climate alter trophic structure in an oceanic ecosystem. Proceedings of the Royal Society of London B 277:1353–1360.
- Simmons, C. M., and S. T. Szedlmayer. 2011. Recruitment of age-0 Gray Triggerfish to benthic structured habitat in the northern Gulf of Mexico. Transactions of the American Fisheries Society 140:14–20.
- Taquet, M., L. Dagorn, J. C. Gaertner, C. Girard, R. Aumerruddy, G. Sancho, and D. Itano. 2007. Behavior of Dolphinfish (*Coryphaena hippurus*) around drifting FADs as observed from automated acoustic receivers. Aquatic Living Resources 20:323–330.
- Taylor, S. M., R. T. Leaf, F. J. Hernandez, J. S. Franks, and E. R. Hoffmayer. 2015. An examination of the diversity and abundance of ichthyoplankton in the Loop Current in the central Gulf of Mexico. Proceedings of the Gulf and Caribbean Fisheries Institute 67:173–185.

- Thompson, N. B. 1999. Characterization of the Dolphinfish (Coryphaenidae) fishery of the United States western North Atlantic Ocean. Scientia Marina 63:421–427.
- Torres-Rojas, Y. E., A. Hernández-Herrera, S. Ortega-García, and M. F. Soto-Jiménez. 2014. Feeding habits variability and trophic position of Dolphinfish in waters south of the Baja California Peninsula, Mexico. Transactions of the American Fisheries Society 143:528–542.
- Tripp-Valdez, A., F. Galván-Magaña, and S. Ortega-García. 2010. Feeding habits of Dolphinfish (*Coryphaena hippurus*) in the southeastern Gulf of California, Mexico. Journal of Applied Ichthyology 26:578–582.
- Tripp-Valdez, A., F. Galván-Magaña, and S. Ortega-García. 2015. Food sources of common Dolphinfish (*Coryphaena hippurus*) based on stomach content and stable isotopes analyses. Journal of the Marine Biological Association of the UK 95:579–591.
- Valdez-Moreno, M., C. Quintal-Lizama, R. Gómez-Lozano, and M. D. C. García-Rivas. 2012. Monitoring an alien invasion: DNA barcoding and the identification of Lionfish and their prey on coral reefs of the Mexican Caribbean. PLoS (Public Library of Science) One [online serial] 7(6): e36636.
- Varghese, S. P., V. S. Somvanshi, M. E. John, and R. S. Dalvi. 2013. Diet and consumption rates of common Dolphinfish, *Coryphaena hippurus*, in the eastern Arabian Sea. Journal of Applied Ichthyology 29:1022–1029.
- Wells, R. J., and J. R. Rooker. 2004a. Spatial and temporal patterns of habitat use by fishes associated with *Sargassum* mats in the northwestern Gulf of Mexico. Bulletin of Marine Science 74:81–99.
- Wells, R. J., and J. R. Rooker. 2004b. Distribution, age, and growth of youngof-the year Greater Amberjack (*Seriola dumerili*) associated with pelagic *Sargassum*. U.S. National Marine Fisheries Service Fishery Bulletin 102:545–554.
- Williams, E. H., and L. Bunkley-Williams. 2009. Checklists of the parasites of dolphin, *Coryphaena hippurus*, and Pompano Dolphin, *C. equiselis* with new records, corrections, and comments on the literature. Reviews in Fisheries Science 18:73–93.
- Witherington, B., S. Hirama, and R. Hardy. 2012. Young sea turtles of the pelagic *Sargassum*-dominated drift community: habitat use, population density, and threats. Marine Ecology Progress Series 463:1–22.
- Young, P. 2014. Age, growth, and migration patterns of Dolpinfish (*Coryphaena hippurus*) in the northwestern Gulf of Mexico. Master's thesis. Texas A&M University, Corpus Christi.